

# MS/MS interpretation in identification of unknowns

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## Outline

- Introduction
- **How** to interpret LC-MS and MS/MS data.
- Identification of some conjugated metabolites.
- Sensitivity enhancement through derivatization
- Dereplication of natural products, sub-structure analysis of unknowns

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Triglycerides

Genistein  
(a plant secondary metabolite)

Taxol

## Small molecules are important!!

- 89% of all known drugs and 50% of all drugs are derived from pre-existing metabolites.
- Small molecules are cofactors and signalling molecules to 1000's of proteins.
- 100,000 (lipidome)

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## Knowing the unknowns

NMR  
(non-destructive,  
lack of sensitivity for detecting  
minor compounds)

LC-MS/MS  
(selective, highly sensitives)

High  
resolution)

MS/MS

public/commercial  
mass spec chemical database

An unknown compound is a small molecule that can reproducibly be detected and quantified in a metabolomics experiment, but whose chemical identification has not been elucidated yet (Krumsiek et al., 2012).

**Unknown unknown- not previously cited (Little et al., 2011)**

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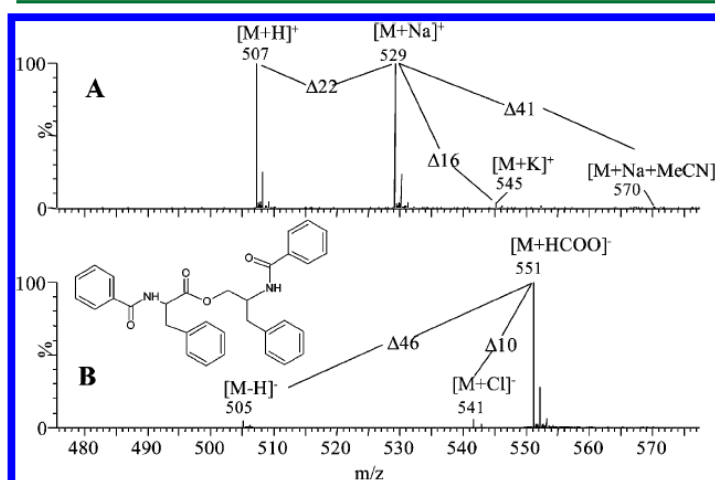
**Keys to identifying unknown structures (putative/definitive) by mass spectrometry**

- Knowing the precursor ion
- Retention time of metabolites in LC
- Accurate mass
- Isotope distribution
- Nitrogen rule
- Fragmentation pattern of a precursor ion
- Product/precursor ion intensity ratio
- Comparison with authentic standards (definitive)

Moco et al. Trends in Analytical Chemistry, 2007

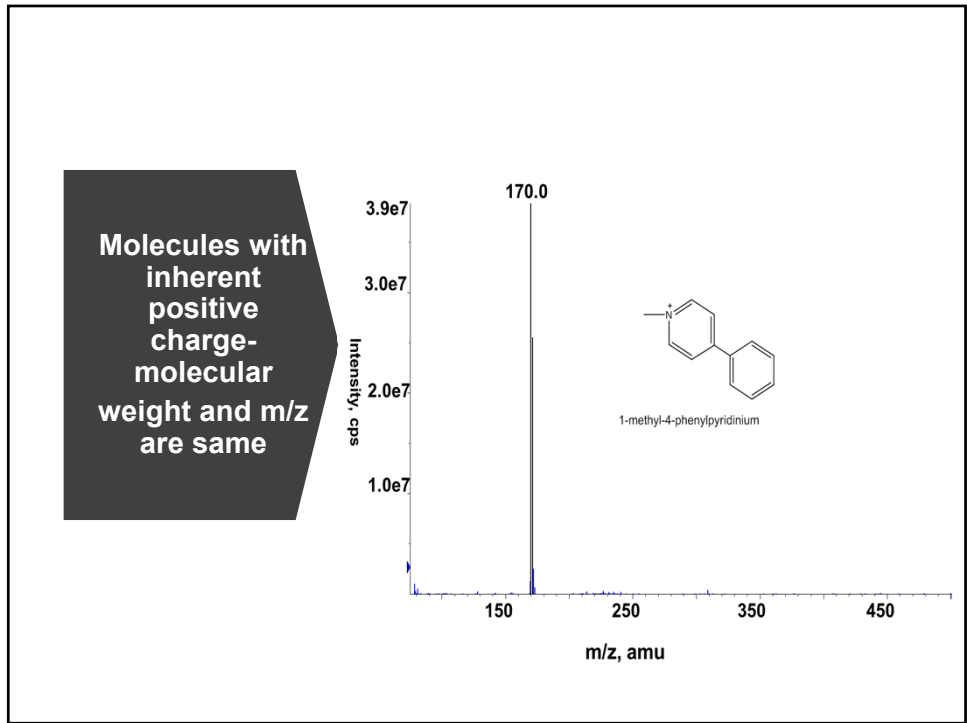
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**Adduct formation in +/-ve ion modes**

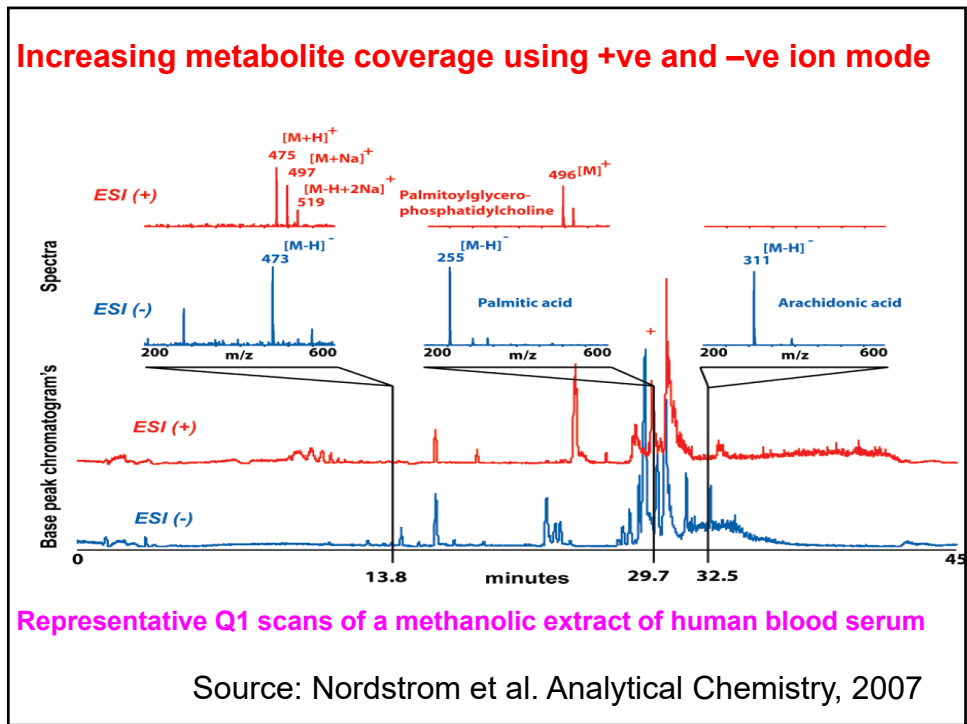


Nielsen et al., J Nat Prod. 2011

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## Isotopic distribution and MS

$1\text{H} = 99.9\%$ ,  $2\text{H} = 0.015\%$

$12\text{C} = 98.9\%$ ,  $13\text{C} = 1.1\%$

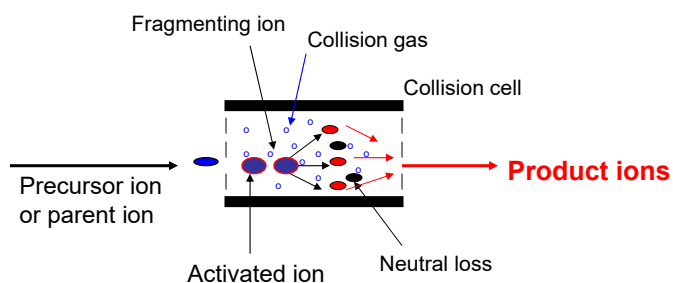
$35\text{Cl} = 75.7\%$ ,  $37\text{Cl} = 24.2\%$

**Monoisotopic mass** - the mass of the most abundant isotope

**Average mass** - the abundance weighted mass of all isotopic components.

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## What is Collision Induced Dissociation (CID) or Collisionally Activated Dissociation (CAD) ?



Schematic of CID fragmentation

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## Applications of MS/MS

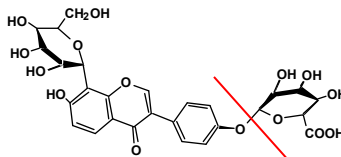
- Pharmaceuticals- Identification and quantification of drug metabolites, PK/PD
- Academic/biotechnology- analysis of protein/peptides, authentication and profiling of chemical components in a crude mixture, substructure analysis of unknown components
- Clinical- eg. neonatal screening, steroids in athletes etc.
- Environment- eg. dioxins in fish..
- Geological- eg. oil compositions...

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## Interpreting MS/MS spectra

- Likely sites of protonation or deprotonation.
- Likely leaving group after cleavage
- Relative abundance of fragment ions
- Neutral molecules are lost

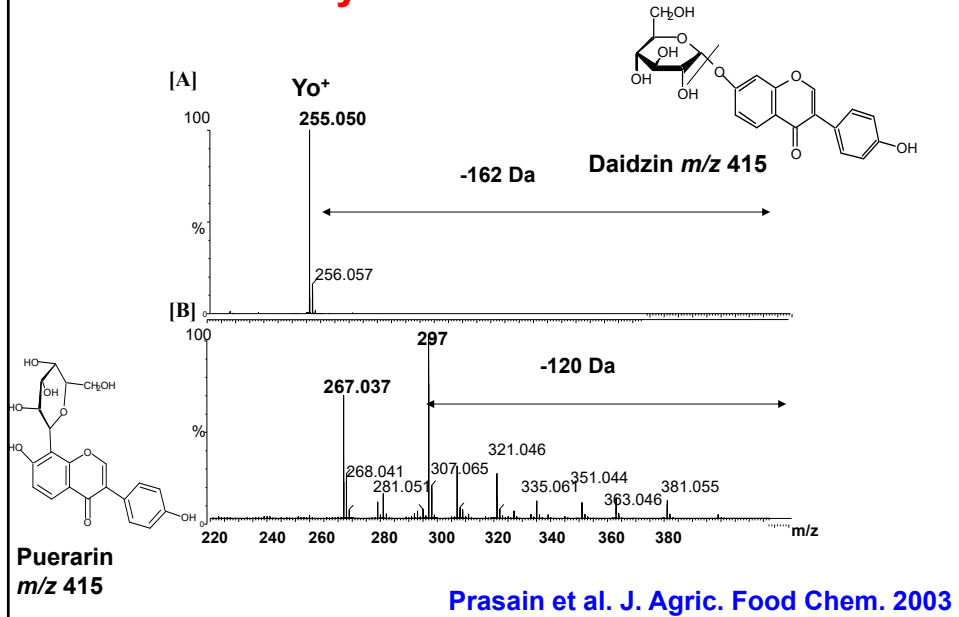
Where are the sites of deprotonation/protonation?  
What is the most likely leaving group in this molecule?



Fragmentation always follows the basic rules of chemistry

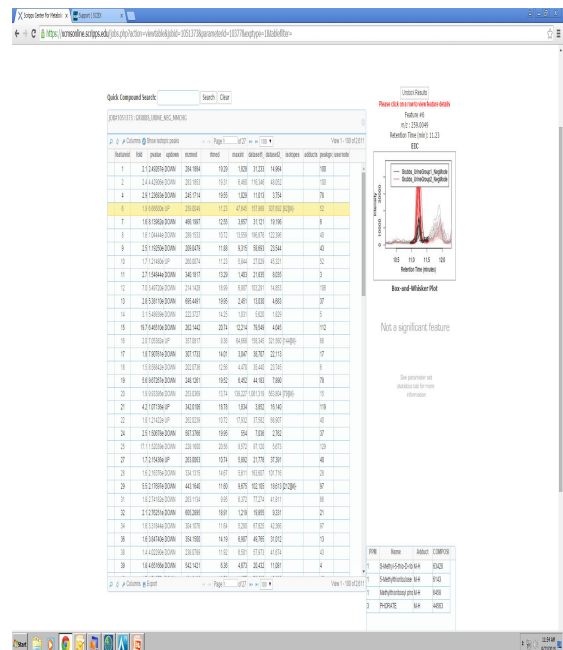
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## O- and C-glucosides fragment differently in ESI-MS/MS



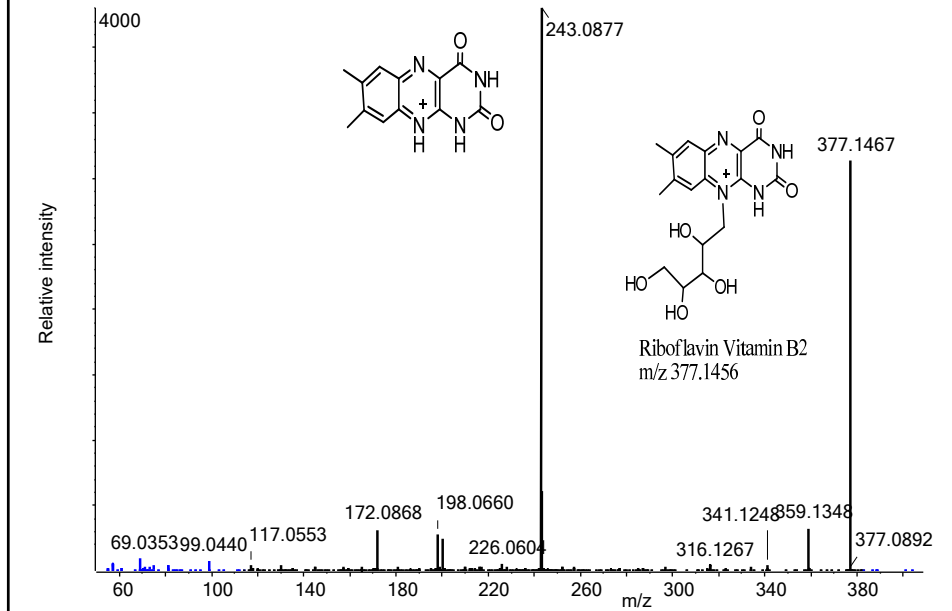
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- XCMSonline screen shot showing list of potential
- metabolites, with p-values and fold changes



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## Detection of Vitamin B2 (riboflavin) as urinary metabolite

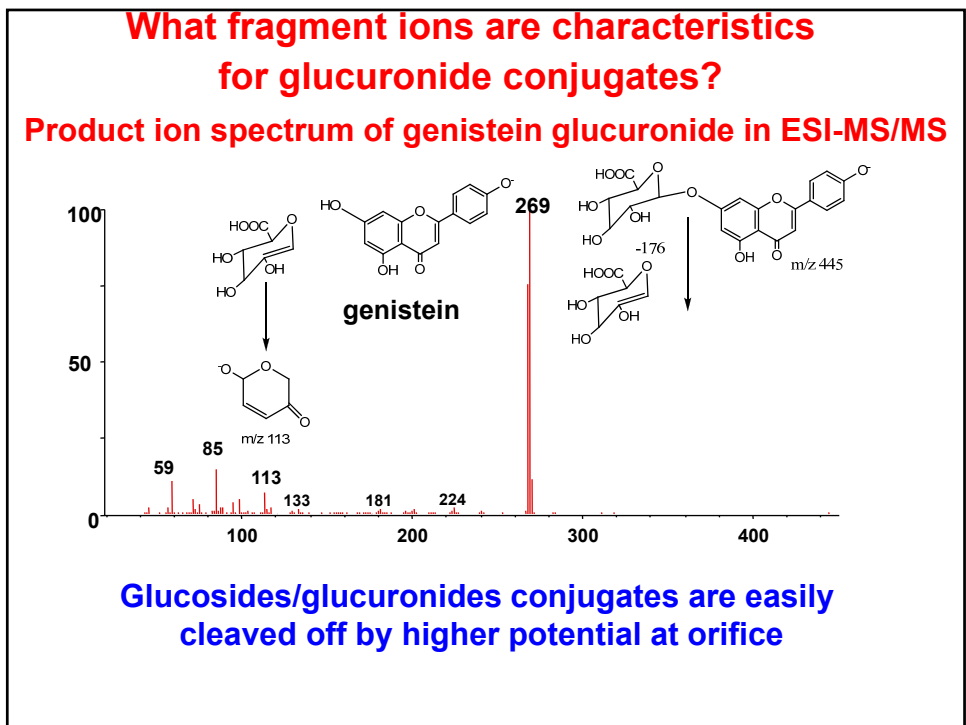


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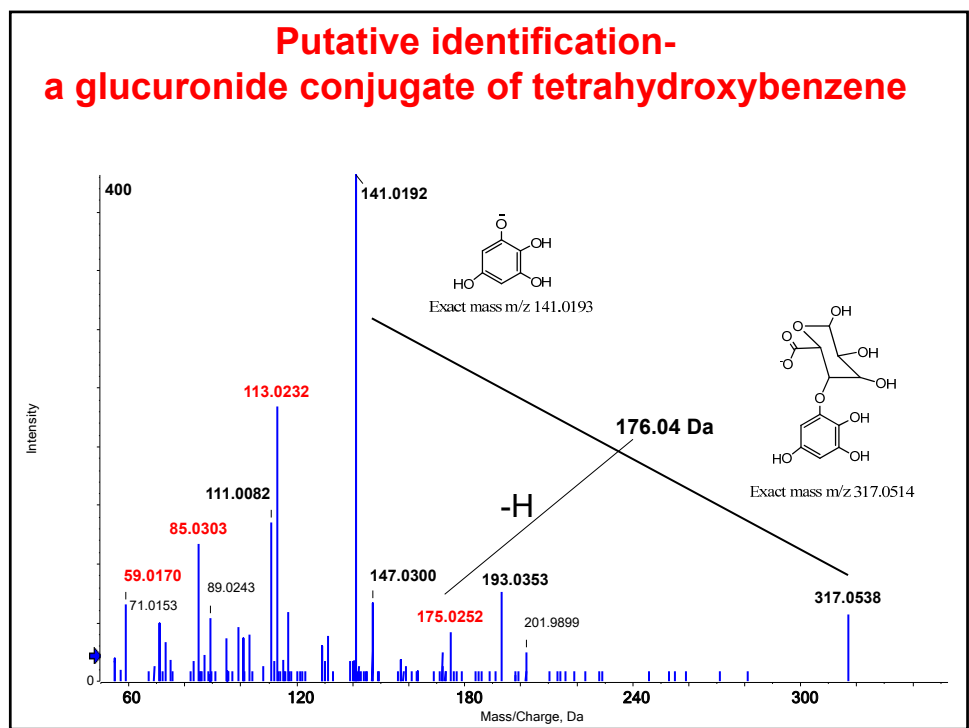
**Ion fragmentation for  
identification of phase II  
drug metabolites  
(glucuronide/sulfate  
conjugates)**

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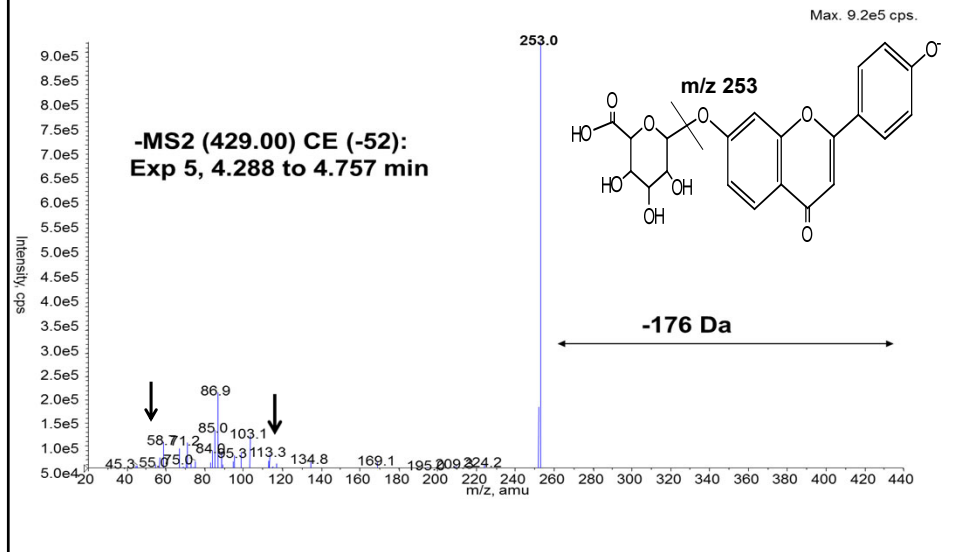


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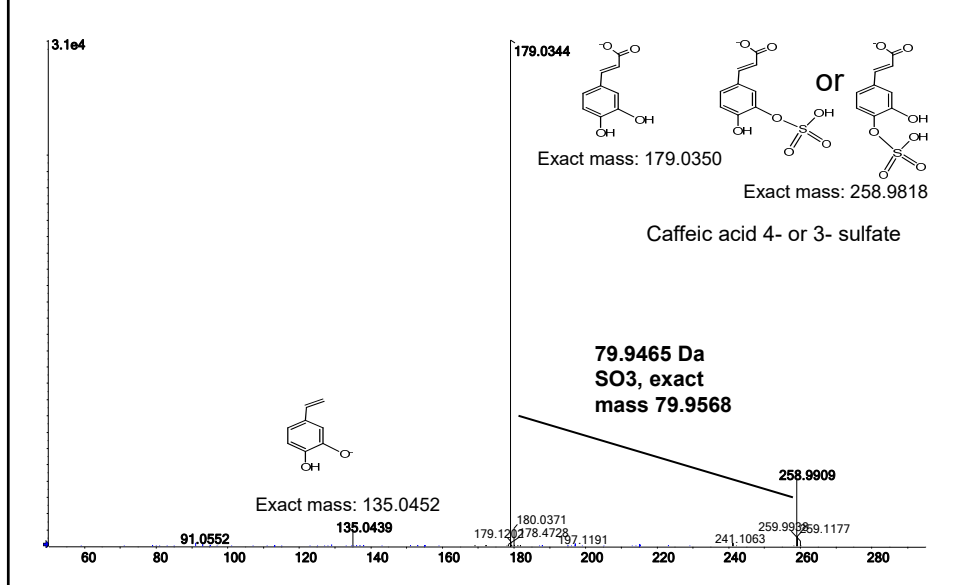
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## MSMS of m/z 429 indicate that it may be daidzein glucuronide



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## High resolution accurate MS/MS help identify sulfated conjugates in unknowns



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## Change in mass is associated with possible metabolic reaction

Metabolic rxn	Change in mass
<b>Methylation</b>	<b>14</b>
<b>Demethylation</b>	<b>-14</b>
<b>Hydroxylation</b>	<b>16</b>
<b>Acetylation</b>	<b>42</b>
<b>Epoxidation</b>	<b>16</b>
<b>Desulfuration</b>	<b>-32</b>
<b>Decarboxylation</b>	<b>-44</b>
<b>Hydration</b>	<b>18</b>
<b>Dehydration</b>	<b>-18</b>

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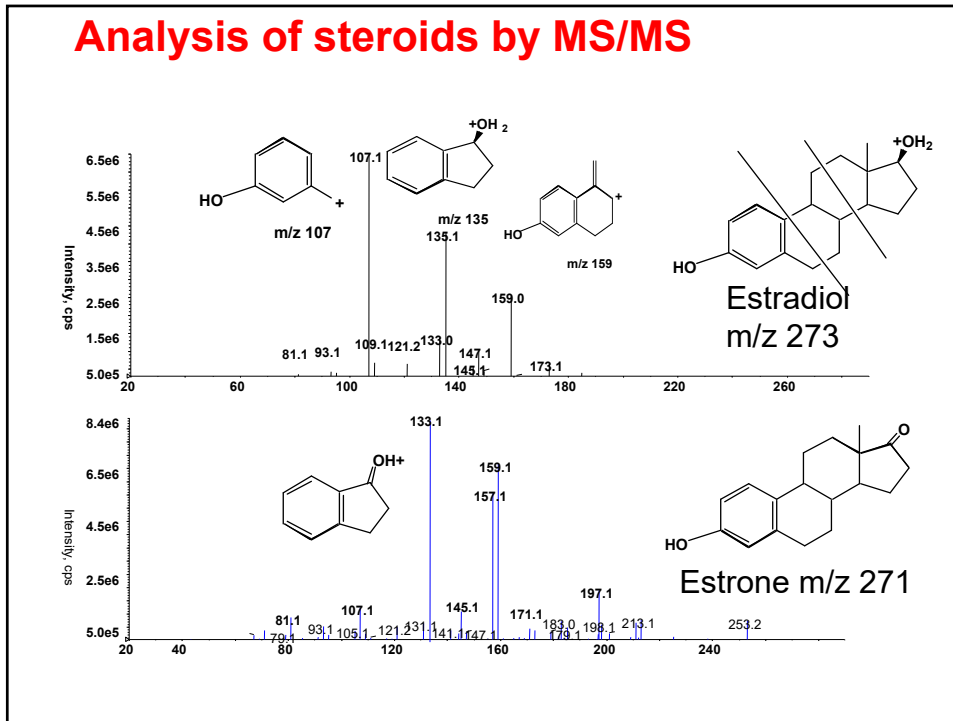
## Characteristic neutral loss and precursor ion scans for conjugated metabolites

Conjugate	Ionization mode	Scan
<b>Glucuronides</b>	<b>pos/neg</b>	<b>NL 176 amu</b>
<b>Hexose sugar</b>	<b>pos/neg</b>	<b>NL 162 amu</b>
<b>Pentose sugar</b>	<b>pos/neg</b>	<b>NL 132 amu</b>
<b>Phenolic sulphate</b>	<b>pos</b>	<b>NL 80 amu</b>
<b>Phosphate</b>	<b>neg</b>	<b>Precursor of m/z 79</b>
<b>Aryl-GSH</b>	<b>pos</b>	<b>NL 275 amu</b>
<b>Aliphatic-GSH</b>	<b>pos</b>	<b>NL 129</b>
<b>taurines</b>	<b>Pos</b>	<b>Precursor of m/z 126</b>
<b>N-acetylcysteins</b>	<b>neg</b>	<b>NL 129 amu</b>

**NL = neutral loss.** **Kostiainen et al., 2003**

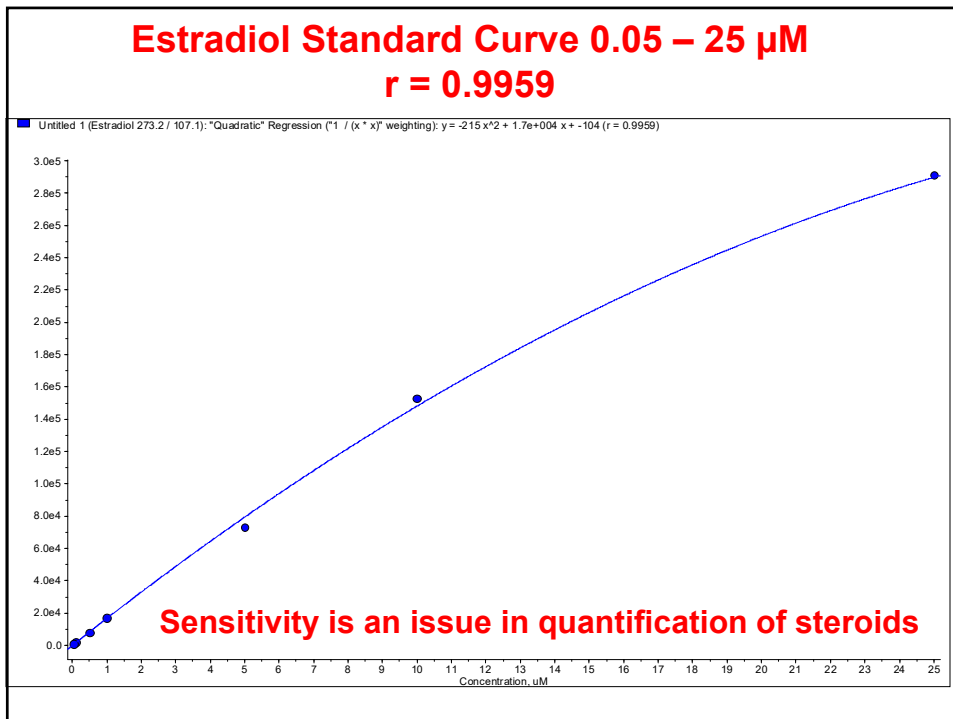
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## Analysis of steroids by MS/MS



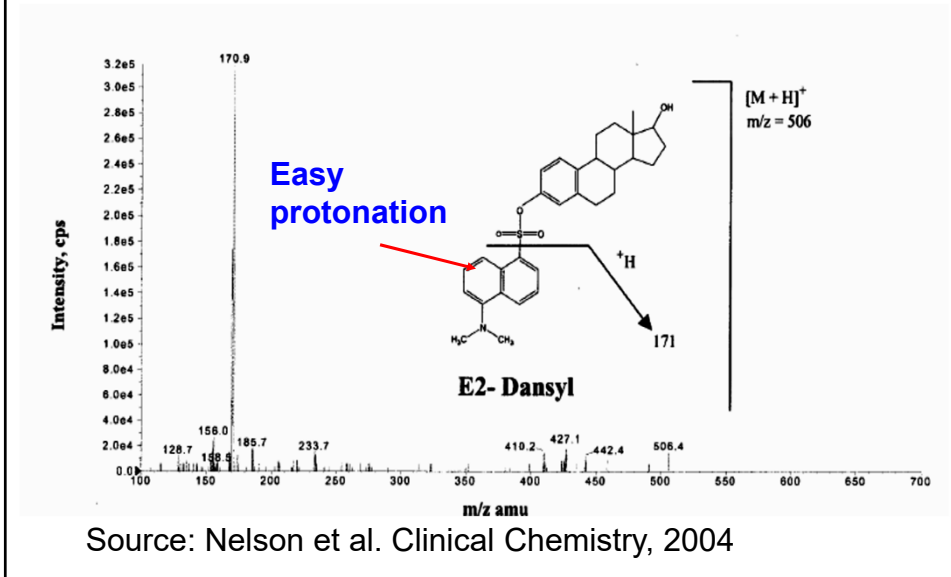
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## Estradiol Standard Curve 0.05 – 25 $\mu\text{M}$ $r = 0.9959$



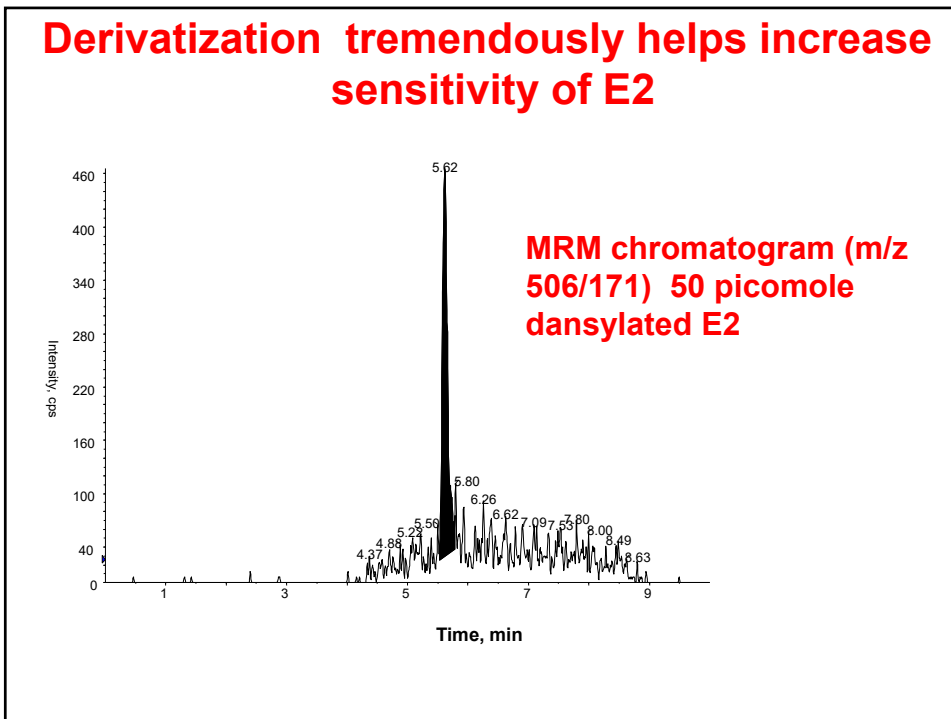
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**Derivatization of estradiol with dansyl chloride leads to the formation of E<sub>2</sub>-dansyl (m/z 506)**



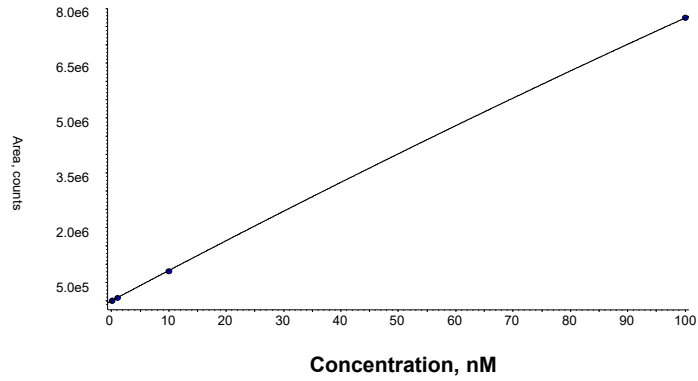
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**Derivatization tremendously helps increase sensitivity of E<sub>2</sub>**



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**Calibration curve for dansylated E2 showing  
linearity from 0.005-100 nM concentration range  
( $r = 0.999$ )**

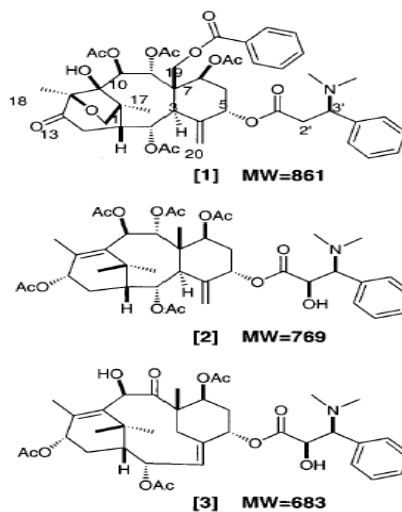


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**Substructure analysis in ESI-MS/MS  
(dereplication and partial identification  
of natural products)**

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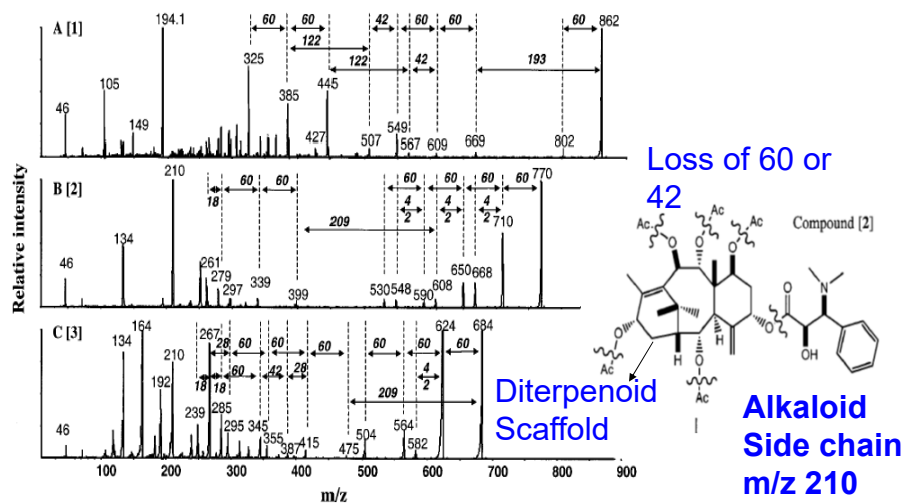
## Fragmentation of basic taxoids from *T. Wallichiana* extract



*Stefanowicz et al. Anal Chem, 2001*

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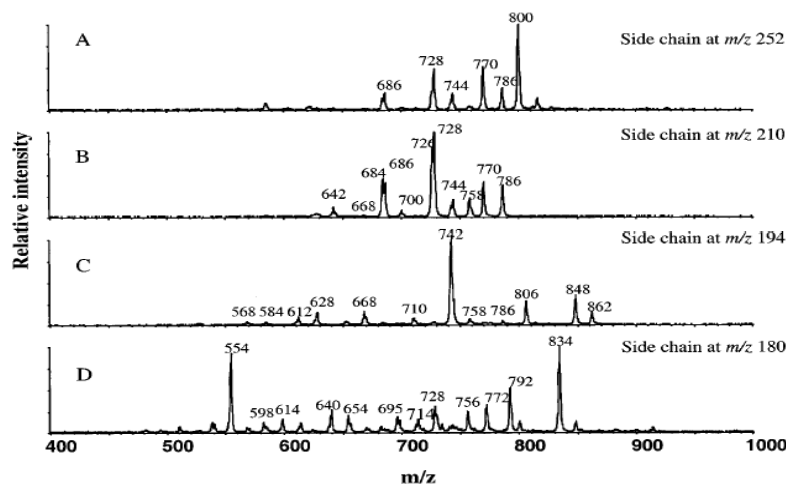
### ESI-MS/MS spectra of taxoids (1-3). Peaks $m/z$ 194 and 210 represent the intact alkaloid side chain.



*Stefanowicz et al. Anal Chem, 2001*

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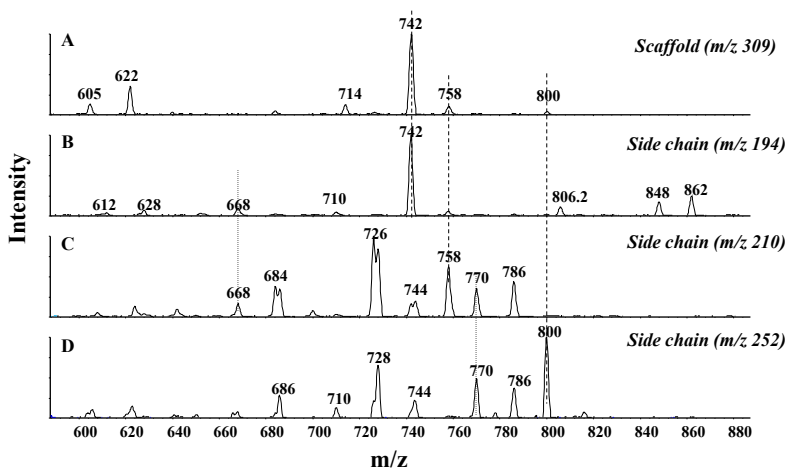
**MS/MS precursor-scan spectra of typical alkaloid side chains to identify the basic taxoids compounds in an ethyl acetate extract of *T. wallichiana*.**



*Stefanowicz et al. Anal Chem, 2001*

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**Comparison of precursor scan spectra obtained from the scaffold  $m/z$  309 and side chain  $m/z$  194, 210 and 252**



**Taxoids with scaffold  $m/z$  309 and alkaloid side chains are shown by dashed lines**

*Stefanowicz et al. Anal Chem, 2001*

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## References

1. **Electrospray Ionization Mass Spectrometry** by Richard B. Cole.
2. **Stefanowicz P, Prasain JK, Yeboah KF, Konishi Y. Detection and partial structure elucidation of basic taxoids from *Taxus wallichiana* by electrospray ionization tandem mass spectrometry. Anal Chem. 2001;73:3583-9.**
3. [Prasain J.K., Wang C.-C., Barnes S. Mass spectrometric analysis of flavonoids in biological samples. \*Free Radical Biology & Medicine\*, 37: 1324-1350, 2004.](#)
4. **William Griffiths. Tandem mass spectrometry in the study of fatty acids, bile acids and steroids. Mass Spectrometry Reviews, 2003;22:81-152.**
5. **Yi et al., Anal Bioanal Chem. 2006.**